

**NANOCRYSTALLINE SYSTEMS FOR PROTECTION,  
DETECTION AND DEMILITARIZATION**

**Final Report**

by

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Research over the last year on grant no R & D 7504-CH-01 from the European Research Office of the U.S. Army has consisted both in the investigation of the photocatalytic activity of mammalian ferritin and in the studies of nanocrystalline oxide films, specifically  $\text{WO}_3$  and  $\text{TiO}_2$ .

The first phase of the project concerns photocatalytic studies on small iron oxide clusters in view of the use of such iron oxide based catalysts in the demineralization of non-stockpile agents. We have initiated work on ferritin which is a very interesting natural protein, carrying in its cavity iron oxide particles with a size of approximately 5 nm. We have discovered that these quantum sized iron oxide particles have a very high activity in promoting the oxidation of a whole series of organics by oxygen, a reaction which is strongly catalysed by the band gap excitation of the semiconductor colloid.

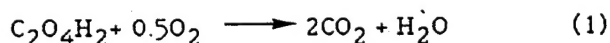
Ferritin, an ubiquitous biological iron-storage protein molecule, consists of 24 symmetrically related protein subunits forming a near-spherical hollow shell, "apoferritin". The central cavity of the apoferritin shell is occupied by an iron core of "ferrihydrite" or  $5\text{Fe}_2\text{O}_3 \cdot 9\text{H}_2\text{O}$  varying in "crystallite" structure (amorphous or crystalline) depending on the source of the ferritin which is widely distributed in nature (e.g., mammalian spleen, liver, heart or bacterial, plant or fungal ferritin.) Through our experimentation it has been shown not only that the iron (III) core can be photochemically reduced (Fe (II)) in presence of electron donors, but likewise that organic substrates such as oxalate and tartrate can be photo-oxidized, the ferrihydrite core acting as a catalyst, with the concomitant reduction of  $\text{O}_2$ . Laser photolysis studies confirmed the reduction of cytochrome C and viologens photosensitized by ferritin via band-gap excitation. (see schemes 1, 2, 3 of Fig. 1).

A schematic representation of the horse spleen ferritin molecule is shown in Fig. 2 a). The source of ferritin used in our experiments was generally Sigma type 1 from horse spleen, in sterile filtered solution in 0.1 M NaCl with a Cd content of less than 1%, although some experimentation using a Fluka source (Biochemika, 50 mg/ml aqueous solution with less than 0.05% Cd, exhibiting less activity) was performed. This lower activity was attributed to a difference in iron core structure or crystallinity. Generally, the ferritin was used as received with no prior treatment in the presence of various chelating agents, such as EDTA. Illumination was carried out employing either a Xenon lamp or in a Hanau suntest apparatus with appropriate filters. Laser photolysis was performed using a doubled frequency ruby laser.

The photoreduction of ferritin (0.25 mg/ml) in presence of 2 mM oxalate, pH 5.5 (50 mM MES buffer) under an argon atmosphere with wavelengths greater than 300 nm is presented in Fig. 3 where after 30 min. illumination practically all of the ferrihydrite is reduced to Fe(II). Re-addition of air results in immediate re-oxidation of Fe(II) to Fe(III). Fig. 4 illustrates the effect of oxalate on this photoreduction. In the absence of oxalate only a very slight reduction of ferritin is witnessed due to electron donation by the protein shell molecules. The effect of pH on this photoreduction reaction, that is, the more

acid the solution, the more rapid the reaction, is demonstrated in Fig. 5. Kinetics of the photoreduction of ferritin in presence of various electron donors such as tartrate, cysteine and ascorbate, as well as the blank (donor electrons from protein shell) are shown in Fig. 6.

The photo-oxidation and resultant full destruction of the organic compounds oxalate and tartrate with concomitant  $\text{CO}_2$  evolution and  $\text{O}_2$  consumption in presence of mammalian ferritin at an initial pH of 3 under suntest lamp conditions was observed. The pH in the unbuffered experiments on oxalic acid varies between 3 and 7 during the course of this reaction.



The photo-oxidative degradation of tartaric acid in presence of ferritin proceeds by the following overall stoichiometry



with the pH varying from the initial value of 3 to pH = 5.8. One could easily imagine the extension of this type of photo-oxidative degradation to various organic CW simulants, such as the organic phosphate containing pesticides.

The direct photo-electron charge transfer from photosensitized ferritin to the molecules cytochrome C and various viologens was demonstrated in laser flash photolysis experiments. These molecules are too large to enter the protein core of the ferritin molecule through one of the channels found in the shell structure and, thus, electron transfer must occur via tunneling to the exterior of the protein molecule. Fig. 2 b) and c) are schematic representations of these photoreductions. Fig. 7 illustrates the absorbance changes as a function of time both under dark and illuminated conditions. Figs. 8 and 9 depict the photoreduction of cytochrome C by ferritin with and without the presence of additional electron donors, or air, and as a function of light intensity. The reaction is greatly enhanced in the presence of tartrate. The more acid pH ranges are optimal for cytochrome C photoreduction as for photoreduction of ferritin itself in presence of oxalate; however, the reaction extent for photoreduction of cytochrome C in presence of tartrate remains unchanged over the pH range 5.5 to 8.5. The photoreduction of dimer viologen (DV) by ferritin in presence of tartrate is shown as well in Fig. 10. A flash photolysis experiment demonstrating the direct electron transfer from ferritin to PVS (propyl viologen sulfonate) in presence of tartrate as well as its blank are depicted in Fig. 11.

We have also engaged studies on nanocrystalline oxide films, as mentioned previously. The goal here is to obtain films that exhibit high photocatalytic activity under visible light. To this end, nanocrystalline  $\text{WO}_3$  films in the micron thickness range and with a roughness factor exceeding 1000 have been produced via a colloidal precursor solution. These films have been subjected to

photoelectrochemical studies using methanol as model substrate for oxidation. Valence band hole transfer to this scavenger leading to complete mineralization was witnessed. For comparison, photoelectrochemical phenol oxidation on sensitizer-derivatized thin film  $\text{TiO}_2$  optically transparent electrodes was also investigated.

The colloidal  $\text{WO}_3$  precursor solution was prepared by first dissolving 3.6 gm of  $\text{H}_2\text{WO}_3$  (tungstic acid) in 50 ml of water to which were added 4 ml of concentrated  $\text{NH}_3$ . This solution was then diluted to 800 ml and boiled for 6 hours. The final solution, a transparent colloidal solution of white tungstic acid, attains  $\text{pH}=3.7$ .

The  $\text{WO}_3$  particulate films are prepared by adding 0.1 ml of 4% PVA (polyvinyl alcohol) to 0.1 ml of the 2 M  $\text{WO}_3$  precursor colloidal solution and diluted to 0.24 ml by adding 0.04 ml  $\text{H}_2\text{O}$ . We then applied 0.1 ml of this mixture to a conducting glass (Nippon Sheet Glass, 10 ohm/ $\square$ , fluorine-doped  $\text{SnO}_2$  glass (TCO)) surface of 3.7  $\text{cm}^2$  (E 12 electrode). The film was initially dried in a stream of hot air for 5 to 10 minutes at approximately 90 °C and then sintered at 500 °C under a stream of  $\text{O}_2$  for 1 hour. The scanning electron microscope images depicted in Fig. 12 illustrate the particulate nature of the porous film (7 $\mu$ ) electrode, specifically E 12, for which the preparation is described above. The white light photocurrent/voltage plots of aqueous and 1.5 M methanol solutions, respectively, employing the E 12,  $\text{WO}_3$  working electrode are illustrated in Figs. 13 and 14, the respective bias necessary to obtain charge separation for the efficient oxidation of  $\text{H}_2\text{O}$  and methanol being demonstrated.

Fig. 15 shows the  $\text{WO}_3$  particulate film action spectrum at  $\text{pH} = 3$  in the photoelectrochemical oxidation of methanol (1.5 M) in aqueous solution (supporting electrolyte 0.1 M  $\text{NaClO}_4$ ). The WE potential is adjusted to 500 mV vs SSCE. The incident monochromatic photon to current conversion yield (IPCE) is plotted as a function of excitation wavelength. The IPCE values were derived from the photocurrents ( $\text{mA}/\text{cm}^2$ ) by means of the equation

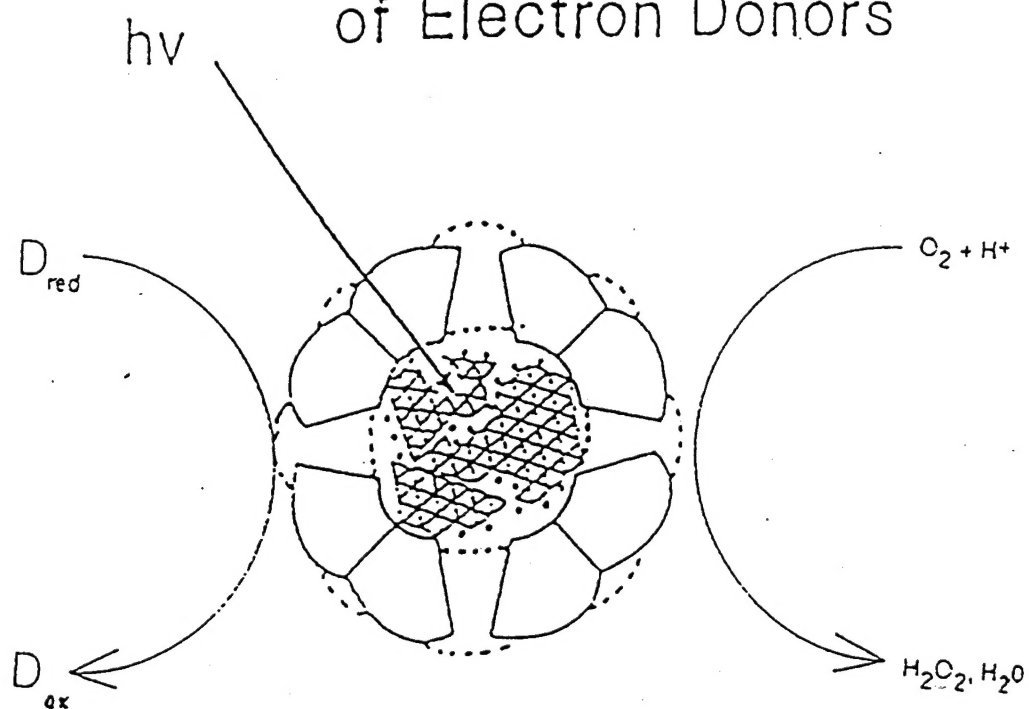
$$\text{IPCE} = 1240 \cdot i_{\text{ph}} / (nm) \cdot P$$

where  $P$  is the incident monochromatic light intensity expressed in  $\text{W}/\text{m}^2$ . The ICPE values increase towards the blue steeply starting from a threshold wavelength at 480 nm. The conversion yield at 400 nm (E12 electrode) attains a value close to 75% in the absence of methanol (Fig. 16) and 115% in the presence of methanol (Fig. 15) indicating that quantitative conversion of incident photons into electric current is taking place. The fact that the yield exceeds 100% is due to the current doubling effect observed with methanol: Hole capture by the latter reagent produces a radical which injects an electron in the conduction band of  $\text{WO}_3$ . From this finding one infers that light induced charge separation is greatly favored in the nanocrystalline oxide films. The decline in efficiency at wavelengths shorter than 400 nm is an artifact due to the glass absorption as can be seen from the inset showing the absorption

spectrum of the film coated  $\text{WO}_3$  electrode. Under polychromatic exposure the experimental photocurrent in presence of methanol was measured as  $3.1 \text{ mA/cm}^2$ , in good agreement with the value calculated of  $3.8 \text{ mA/cm}^2$  from the overlap of the photocurrent action spectrum in Fig. 15 with the standard solar AM 1.5 emission spectrum (see Table 1). In the action spectrum of the  $\text{WO}_3$  electrode we would like to stress the long visible wavelength tail which is not apparent in the action spectrum of the photoelectrochemical oxidation by nanocrystalline  $\text{TiO}_2$ . Fig. 17 illustrates the photocurrent action spectrum of phenol oxidation using a sensitized  $\text{TiO}_2$  particulate film as the WE for comparison with the  $\text{WO}_3$  action spectrum of methanol oxidation. In this case the electrode is a particulate film of  $\text{TiO}_2$  derivatized by the ruthenium complex  $\{\text{Ru}(4,4'-2,2'\text{-bipy})(4'\text{-PO}_3\text{H-terpy})(\text{NCS})\}$ , where  $(4'\text{-PO}_3\text{H-terpy})$  is the novel ligand 2,2':6',2''-terpyridine-4'-phosphonic acid. The supporting electrolyte in this experiment is  $0.01 \text{ M NaClO}_4$ . The second curve depicted is the previous  $\text{RuL}_3$  (ruthenium (2,2')-bipyridyl-(4,4')-dicarboxylic acid) derivatized  $\text{TiO}_2$  film shown for comparison. The phosphonate group of this ligand strongly enhances adsorption onto the  $\text{TiO}_2$  surface and provides sufficient electronic coupling with the oxide to achieve efficient light-induced charge separation. These experiments were carried out in an electrochemical cell equipped with a quartz window, irradiated from the glass side. The source of irradiation was a Xe lamp,  $100 \text{ W/m}^2$  simulated sunlight with a  $470 \text{ nm}$  cut-off filter. Under these conditions any photocurrent is solely attributed to sensitisation. The photocurrent action spectrum indicates a maximum incident photon flux to electron flow conversion efficiency of 18%, compared to 13% for the  $\text{RuL}_3$  sensitized film, at  $480 \text{ nm}$ . The photocurrent calculated for the overlap integral of the action spectra with the AM 1.5 solar emission corresponds to  $1.51 \text{ mA/cm}^2$  for the phosphonated complex and  $0.77 \text{ mA/cm}^2$  for the carboxylated complex, respectively. Thus the new sensitizer based on a phosphonated bipyridyl ligand exhibits a greatly improved visible light response apart from being more strongly adsorbed to  $\text{TiO}_2$  films as compared to the previously employed  $\text{RuL}_3$  dye.

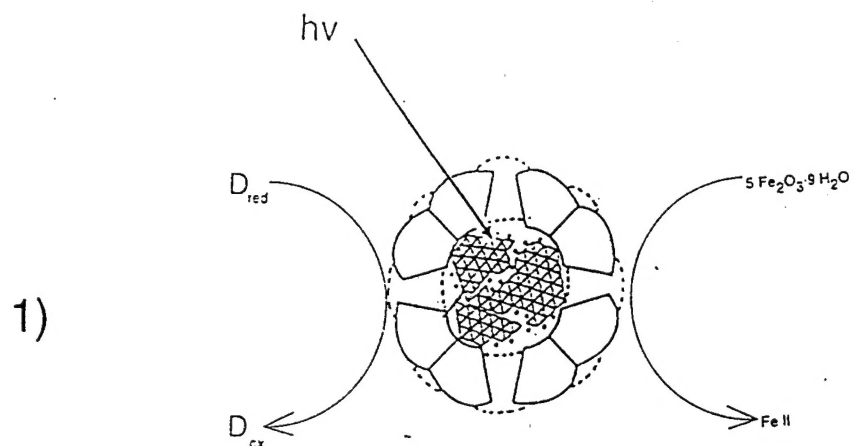
Studies on the reproducibility and efficiency of these nanocrystalline  $\text{WO}_3$  thin film electrodes have advanced greatly. Visible light photoelectrochemical degradation of the simulant compound, 4-nitrophenyldiethylphosphate, Paraoxon, is proving to be quite promising, and will be addressed in a forthcoming report.

# Photoreduction of Ferritin in the Presence of Electron Donors

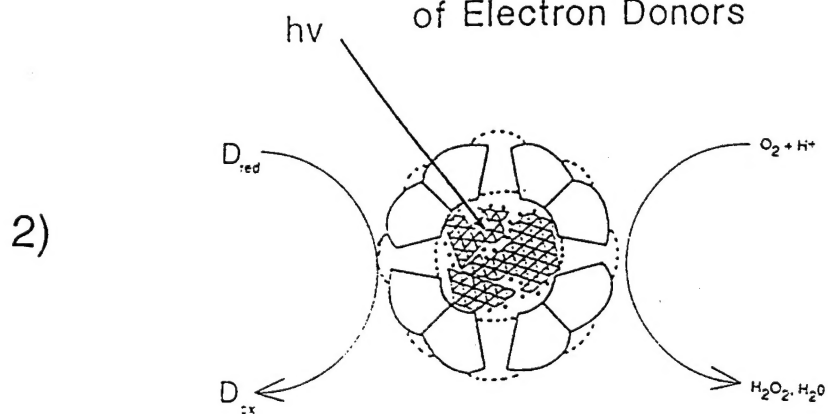


# FIGURE 1

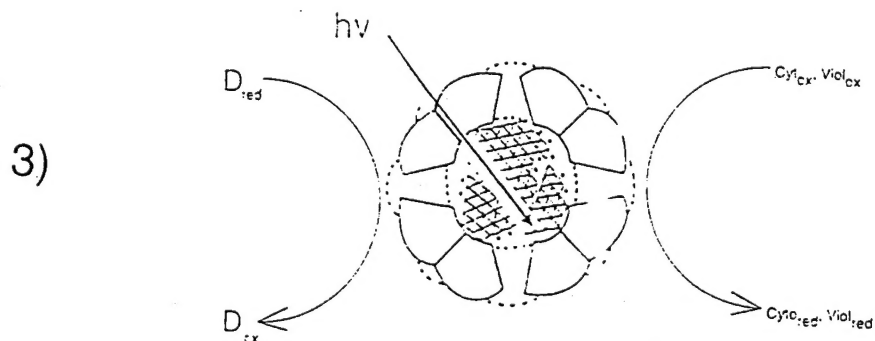
## The Photochemical Activity of Mammalian Ferritin



### Photoreduction of Ferritin in the Presence of Electron Donors



### Photooxidation of the Organic Substrates in the Presence of Ferritin



Reduction of Cytochrome c and Viologens photosensitized by Ferritin

## FIGURE 2

Schematic representation of the horse spleen ferritin molecule

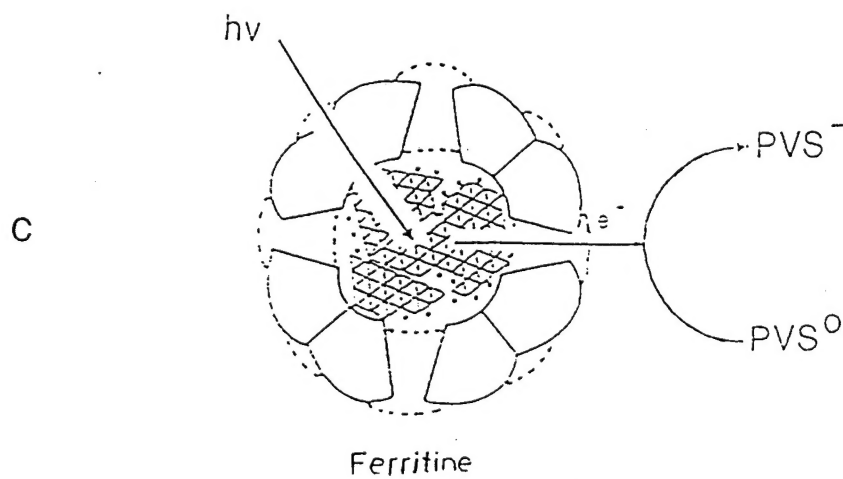
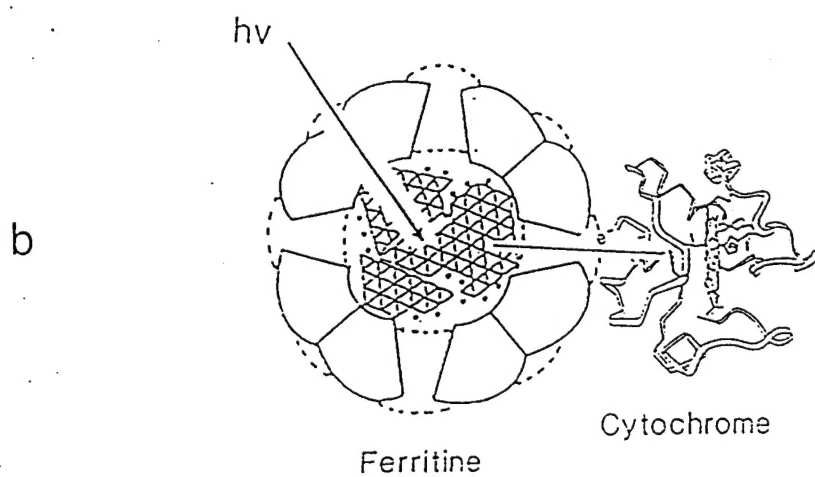
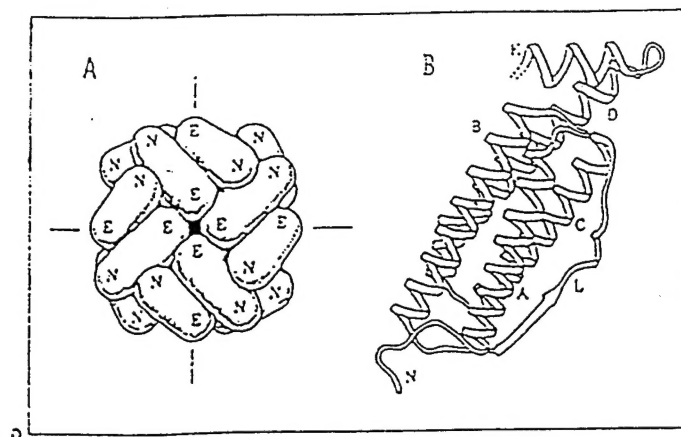
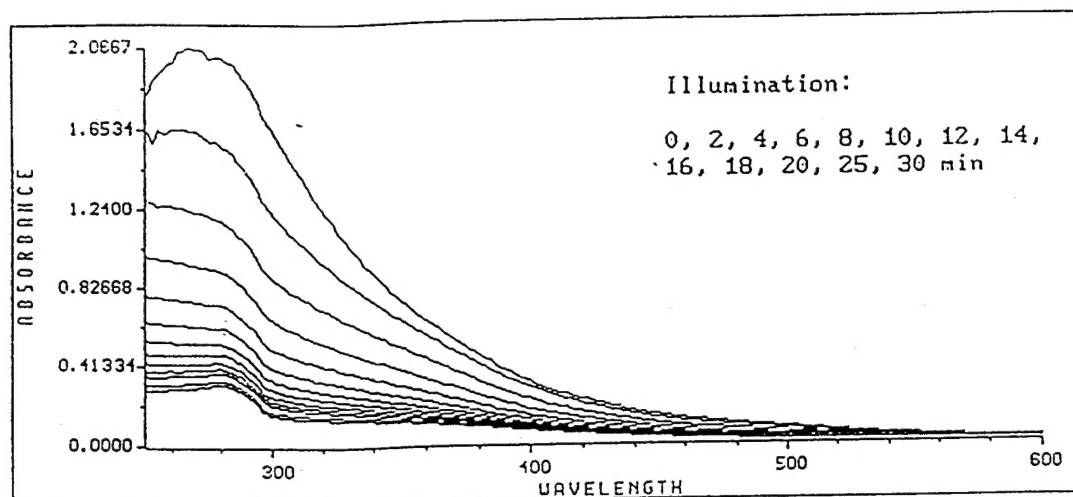




FIGURE 3

Photoreduction of Ferritin  
in the Presence of Oxalate



oxalate, 0.018M, pH 5.0, L=300 nm

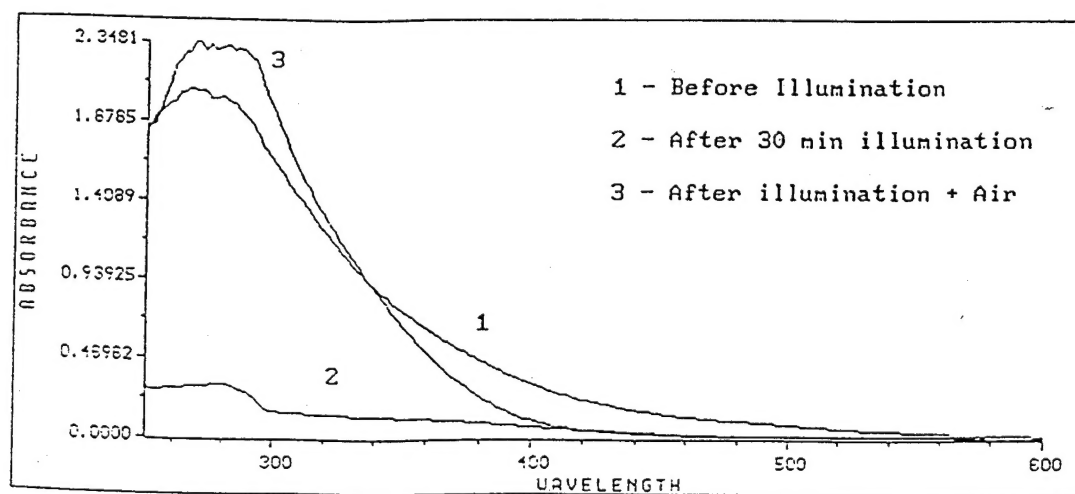
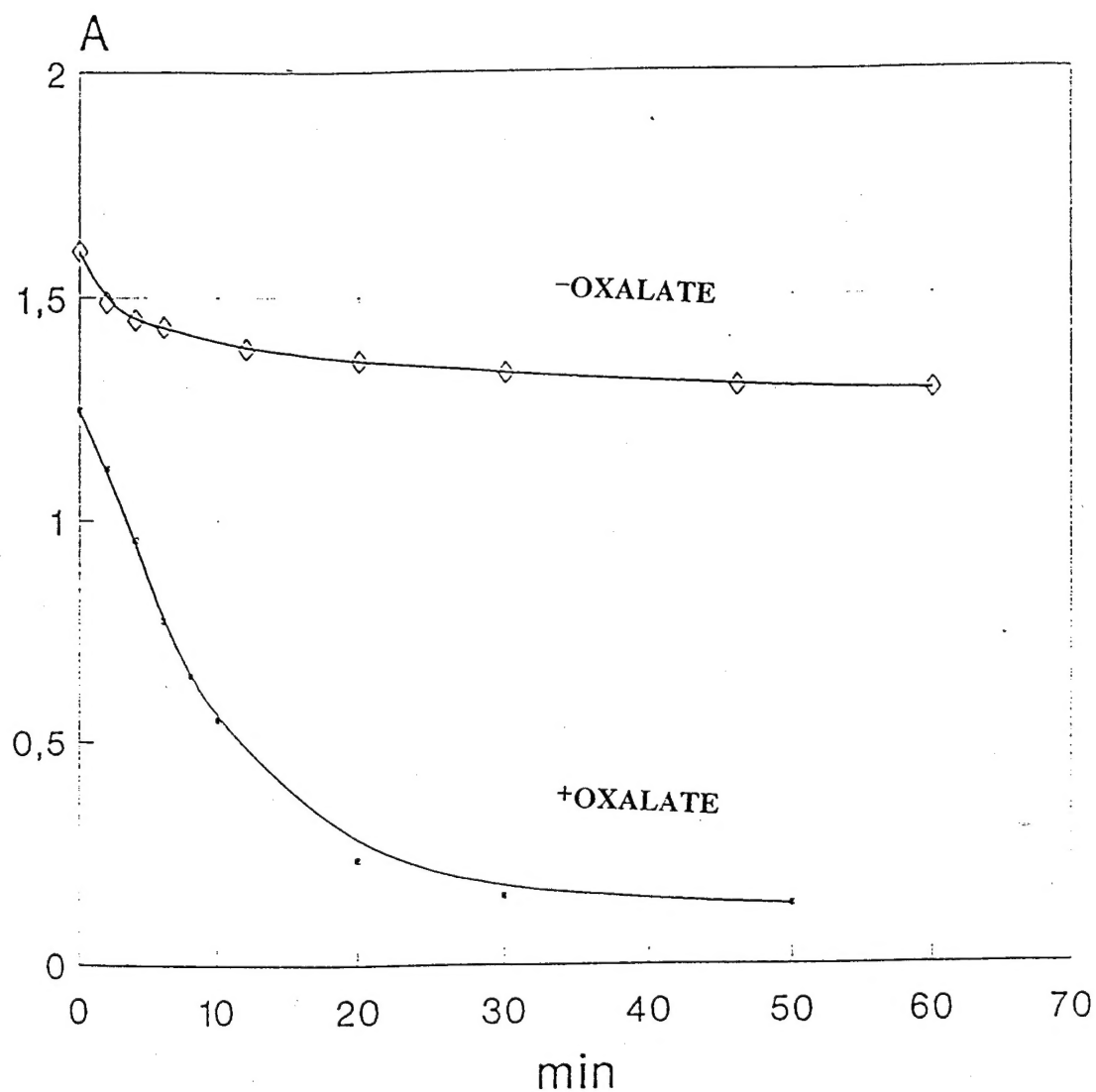


FIGURE 4—

# Photoreduction of Ferritin in the Presence of Oxalate. Kinetics.

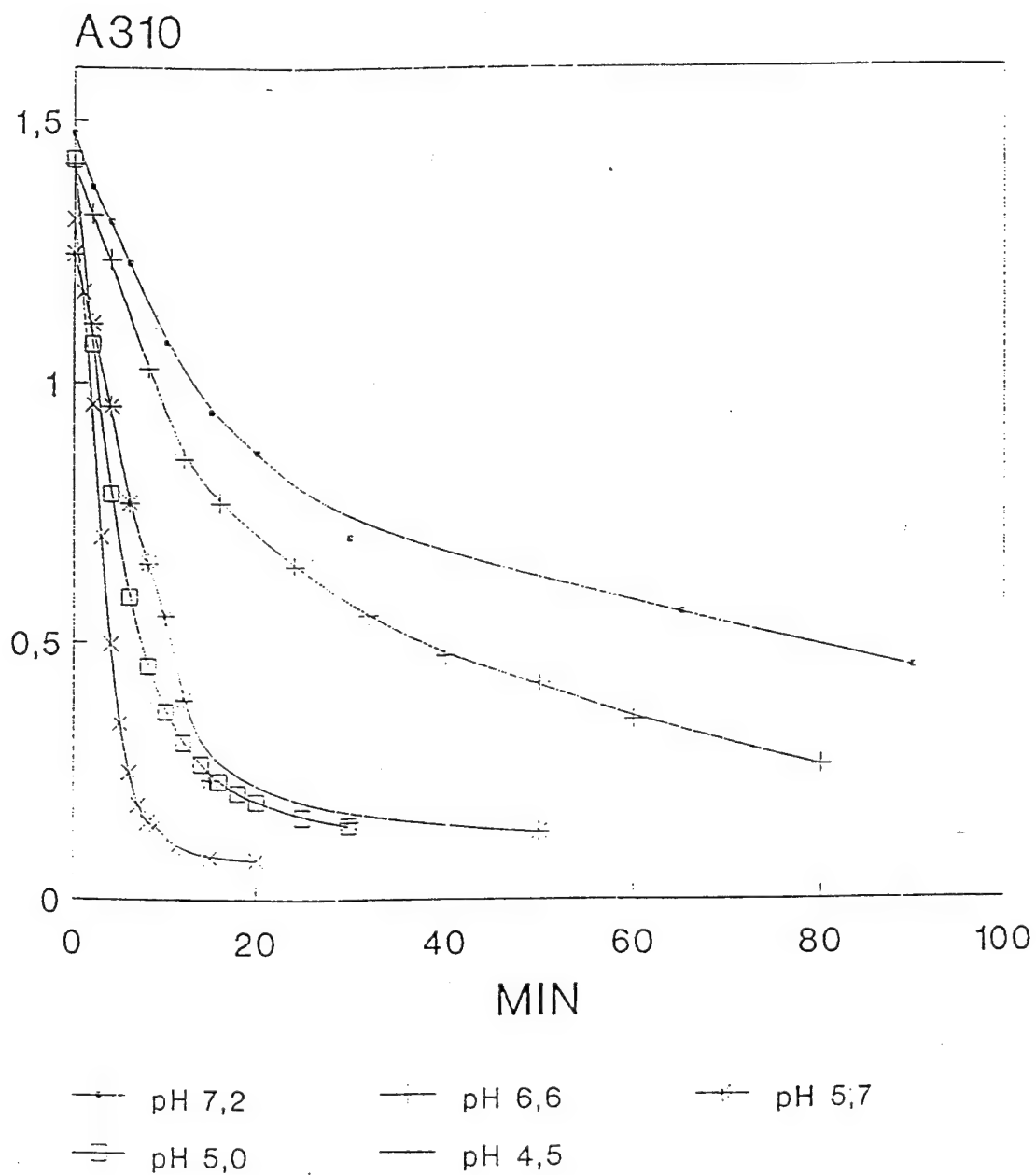


—•— Oxalate, 310 nm      —◇— - Oxalate,  $L > 300\text{nm}$

0.25mg/ml Ferritin, 20mM Oxalate,  
pH 5.5, 50 mM MES, Light  $> 300\text{nm}, > 460\text{nm}$

**FIGURE 5**

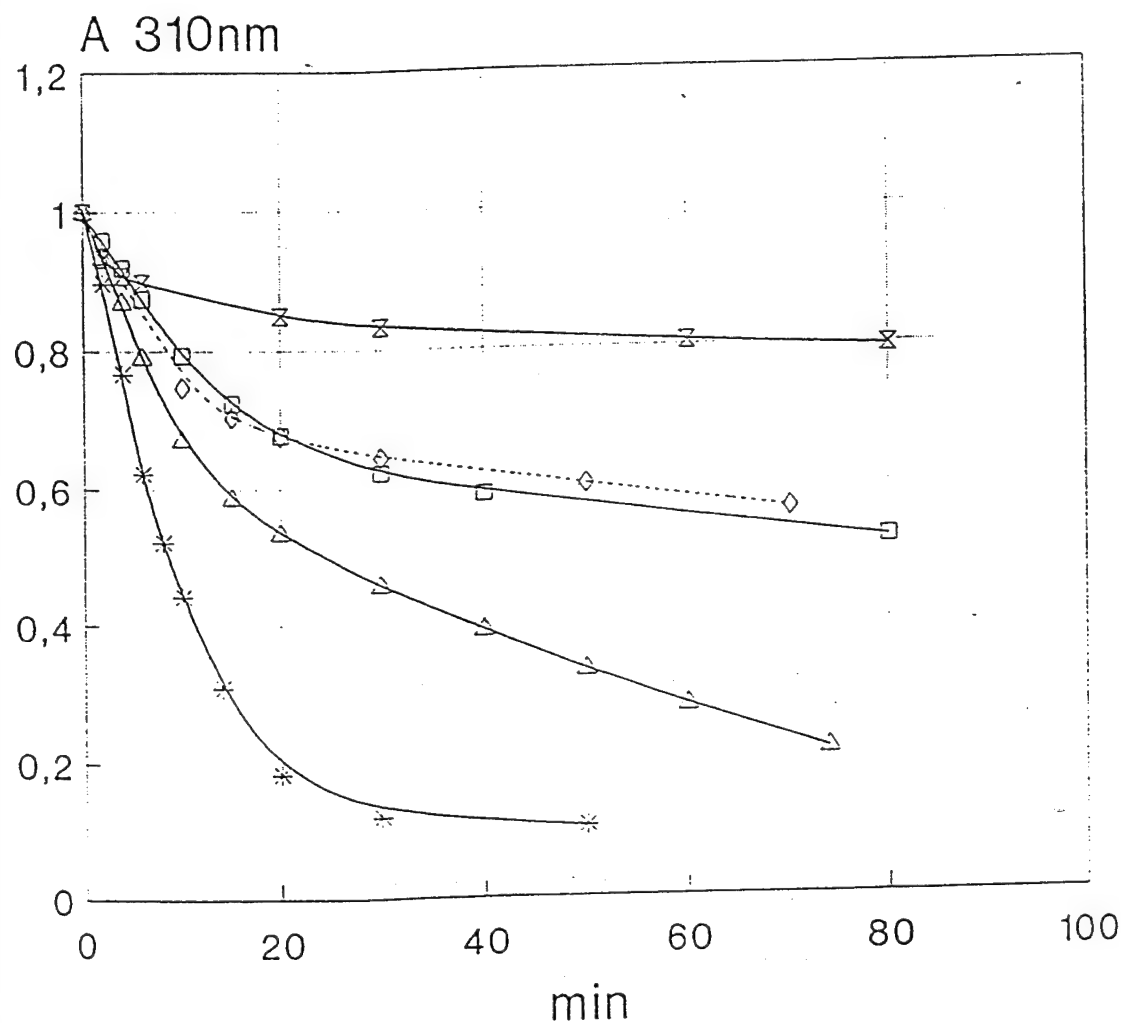
Photoreduction of Ferritin in the  
Presence of Oxalate as a Function of pH



Oxalate 20mM, Fer 0,5mM;  $\lambda > 300\text{nm}$

FIGURE 6

# Photoreduction of Ferritin Kinetics



—\*— Oxalate/      —□— Cysteine/      -◇- Ascorbate/  
 —△— Tartrate/      —— - Donor electron/

0.25mg/ml Ferritin, 20mM donor electron,  
pH 5.5, 50 mM MES, Light > 300nm

FIGURE 7

Photoreduction of Cytochrome by Ferritin

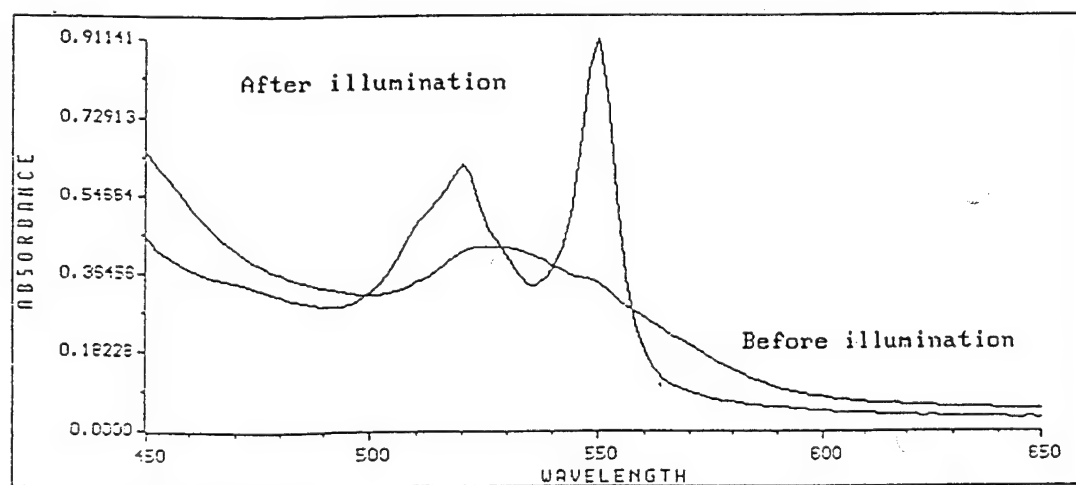
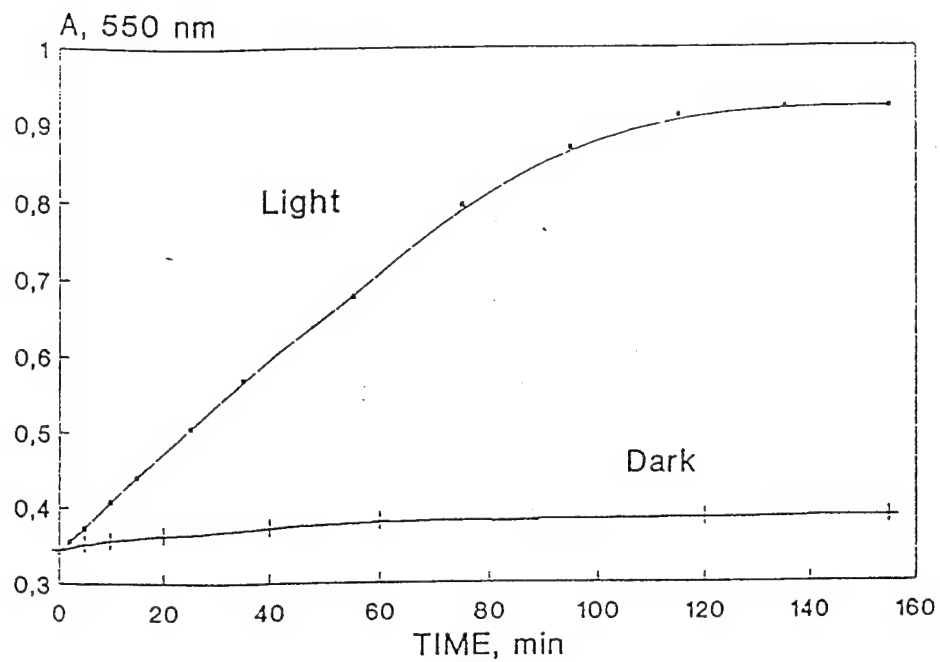
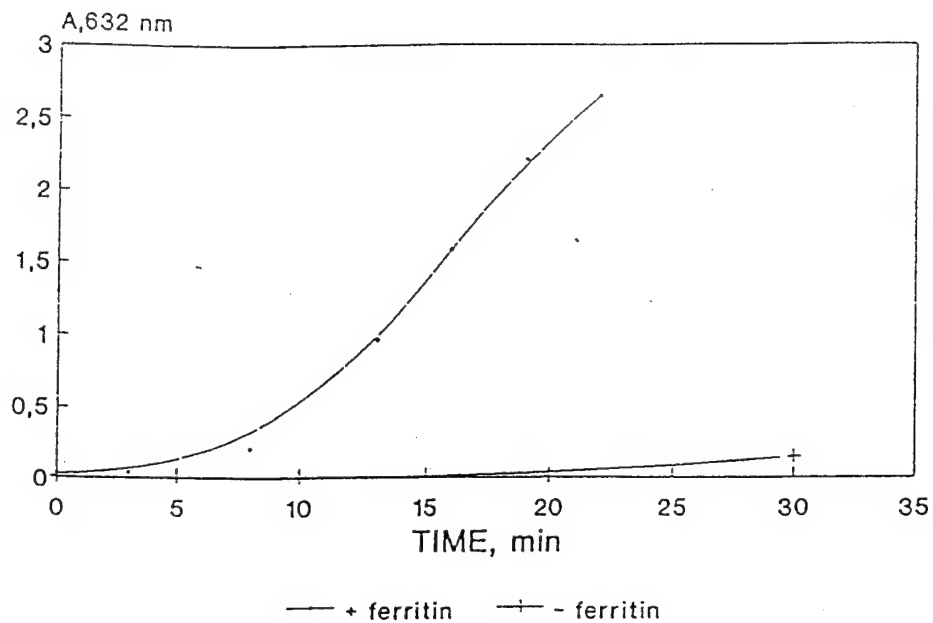
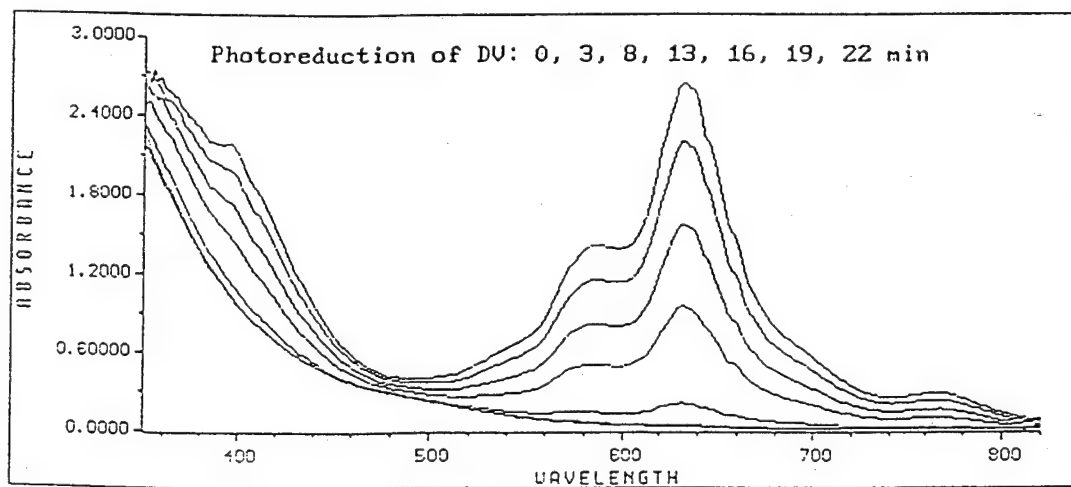


FIGURE 8

## Photoreduction of DV by Ferritin

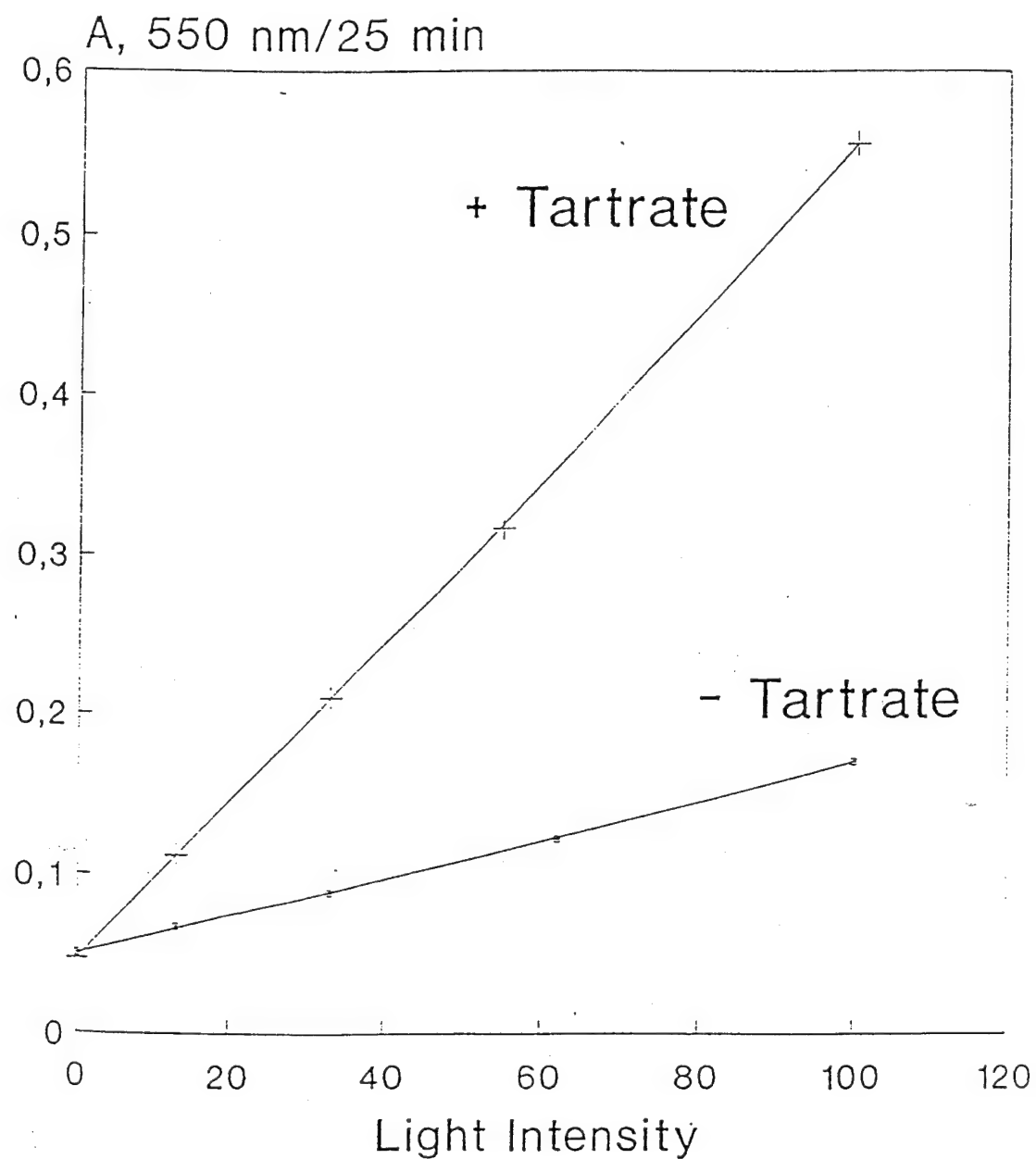


tartrate 0.1M,  $\lambda > 440\text{nm}$ , argon



**FIGURE 9**

Photoreduction of Cytochrome by Ferritin  
as a Function of Light Intensity



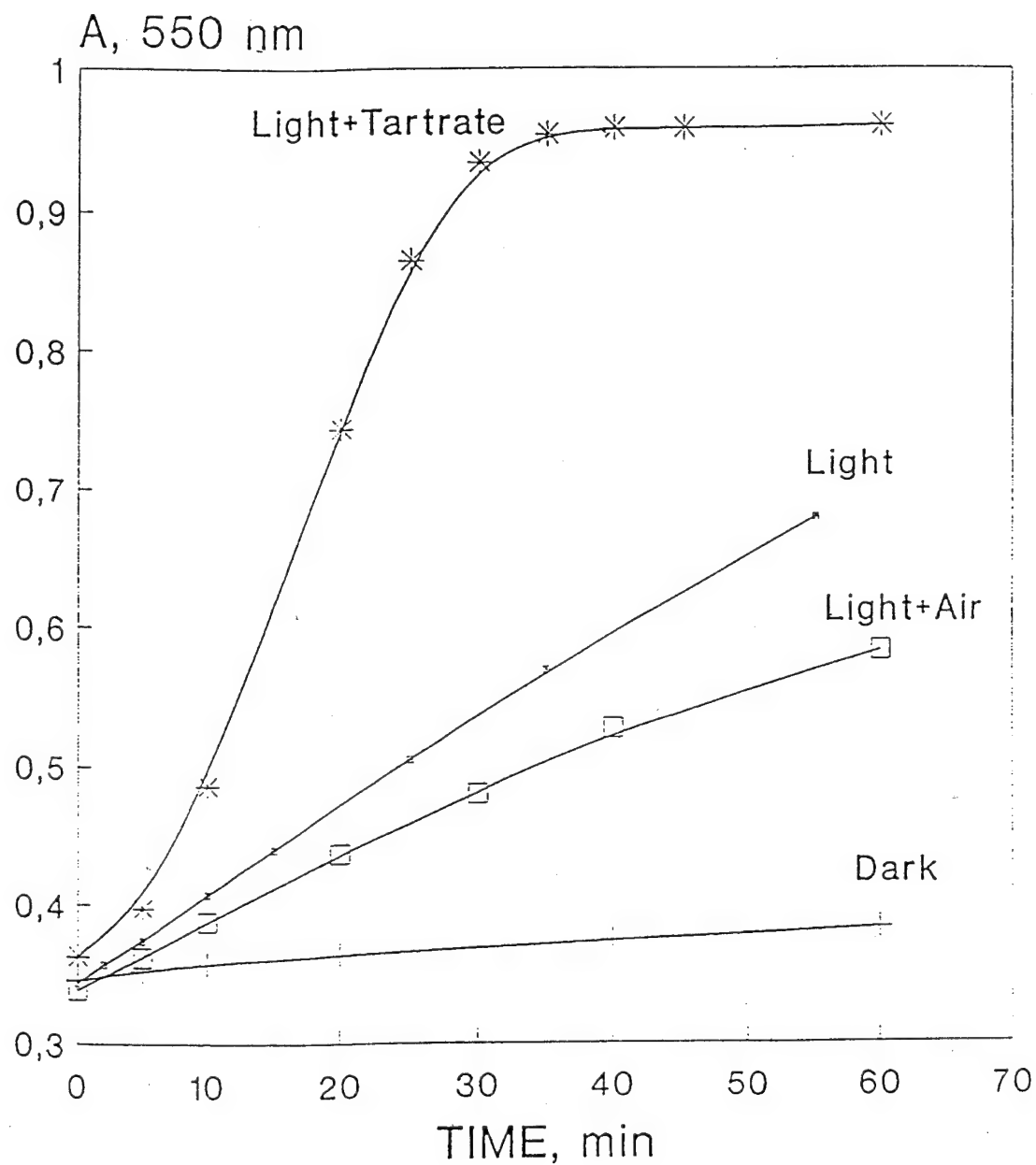
Cytochrome 32mkM. Ferritin 0.5 mg/ml

TES 0.05M, pH 7.0, Light > 440nm

Tartrate 0.02M

FIGURE 10

# Photoreduction of Cytochrome by Ferritin

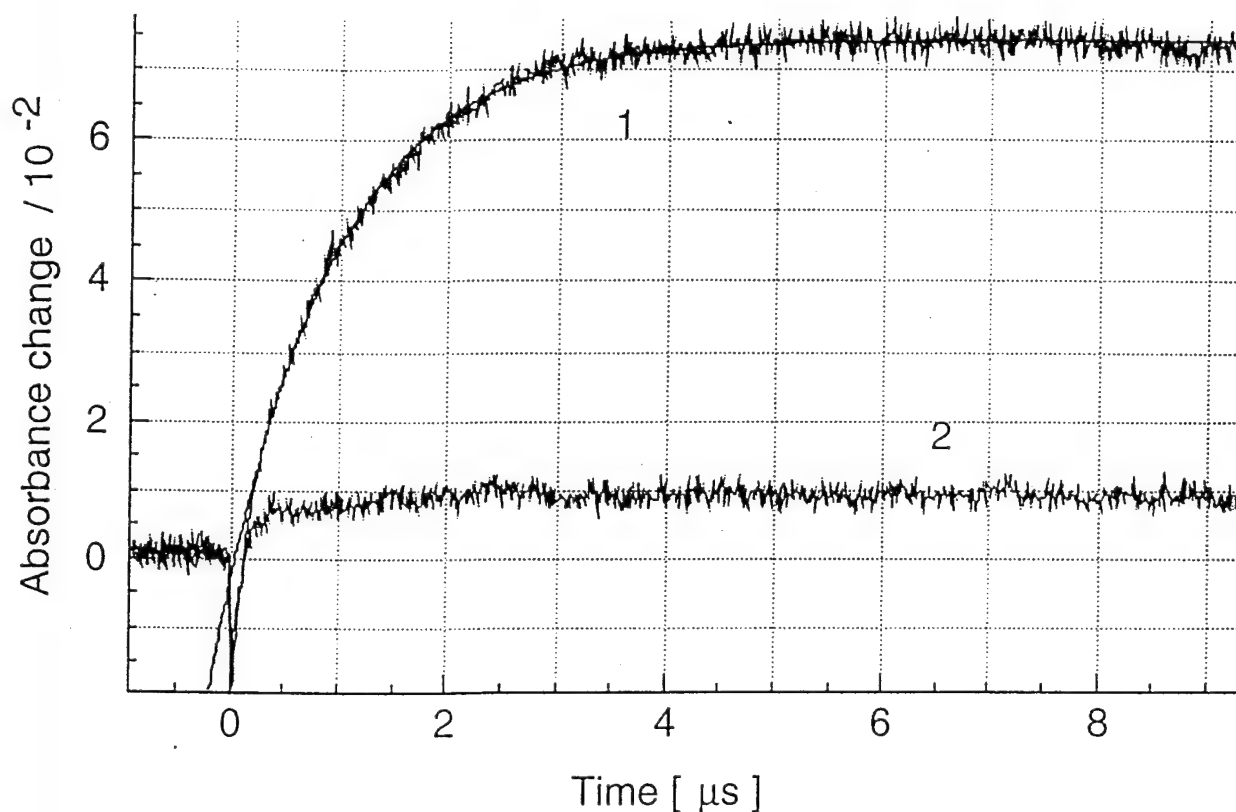


C 32mM, F 1mM; TES 0.05M, pH 7,0;  
0.01M Tartrate, Light >440nm



FIGURE 11

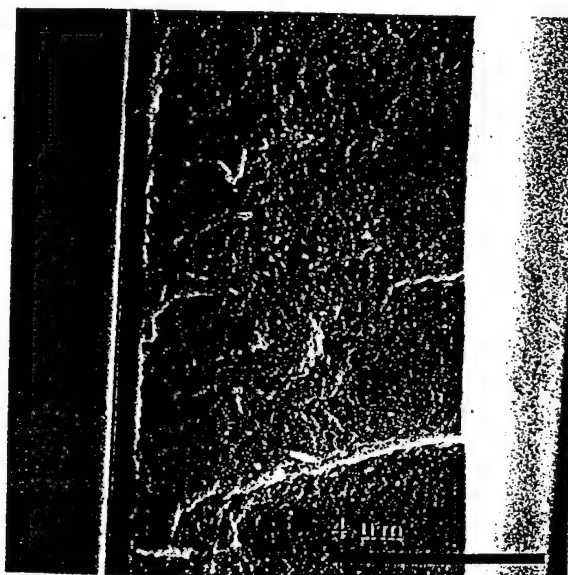
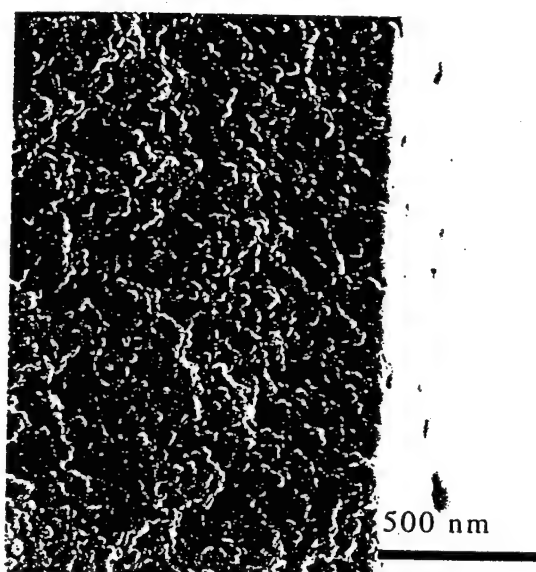
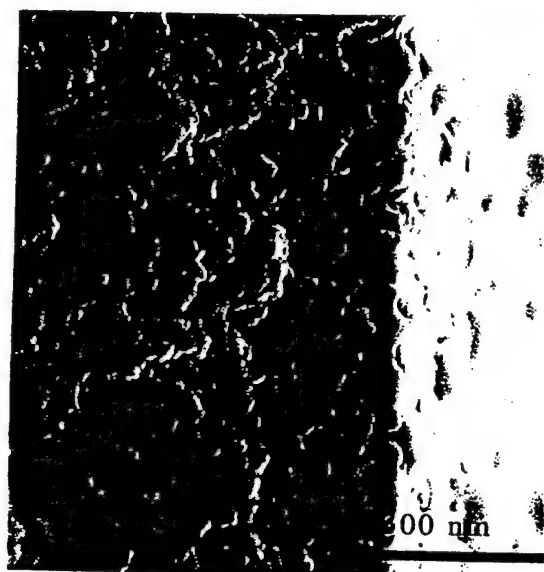
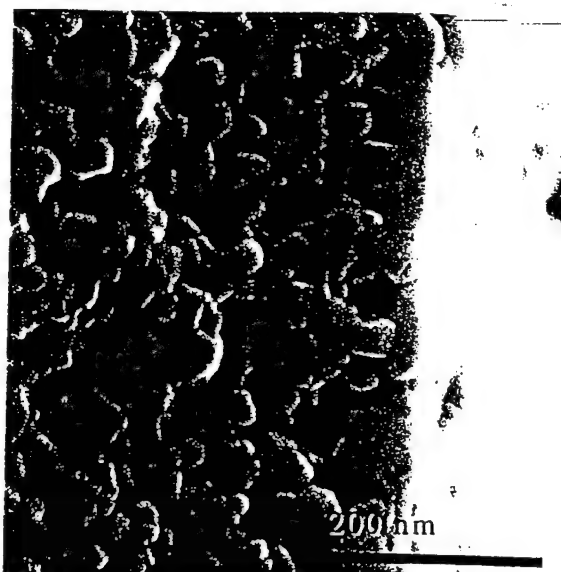
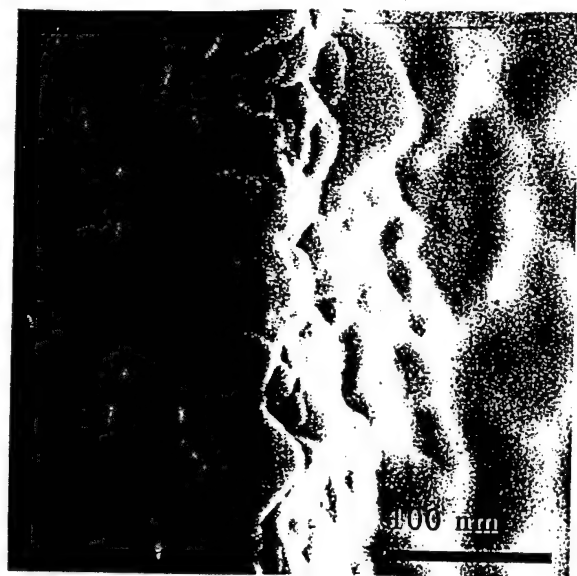
LASER PHOTOLYSIS OF FERRITIN/PROPYLVILOGEN  
SULFONATE AQUEOUS SOLUTIONS



1) Ferritin (horse spleen) 0.11 mg/ml =  $2 \cdot 10^{-7}$  M, Fe(III) in Ferritin =  $1.64 \cdot 10^{-4}$  M,,  
PVS =  $10^{-3}$  M, tartrate =  $10^{-1}$  M, CAPS buffer pH=10, Ar bubbled,  $\lambda_{\text{ext}}=347$  nm,  $\lambda_{\text{obs}}=602$  nm.

2) blank, all conditions identical except no ferritin added

FIGURE 12



SCANNING ELECTRON MICROSCOPE IMAGES OF  
NANOCRYSTALLINE FILM EDGES IN  $\text{WO}_3$ , PREPARATION E12,  
WORKING ELECTRODE

WHITE LIGHT PHOTOCURRENT/VOLTAGE PLOT  
FOR WATER OXIDATION USING WO<sub>3</sub> (E12) WE

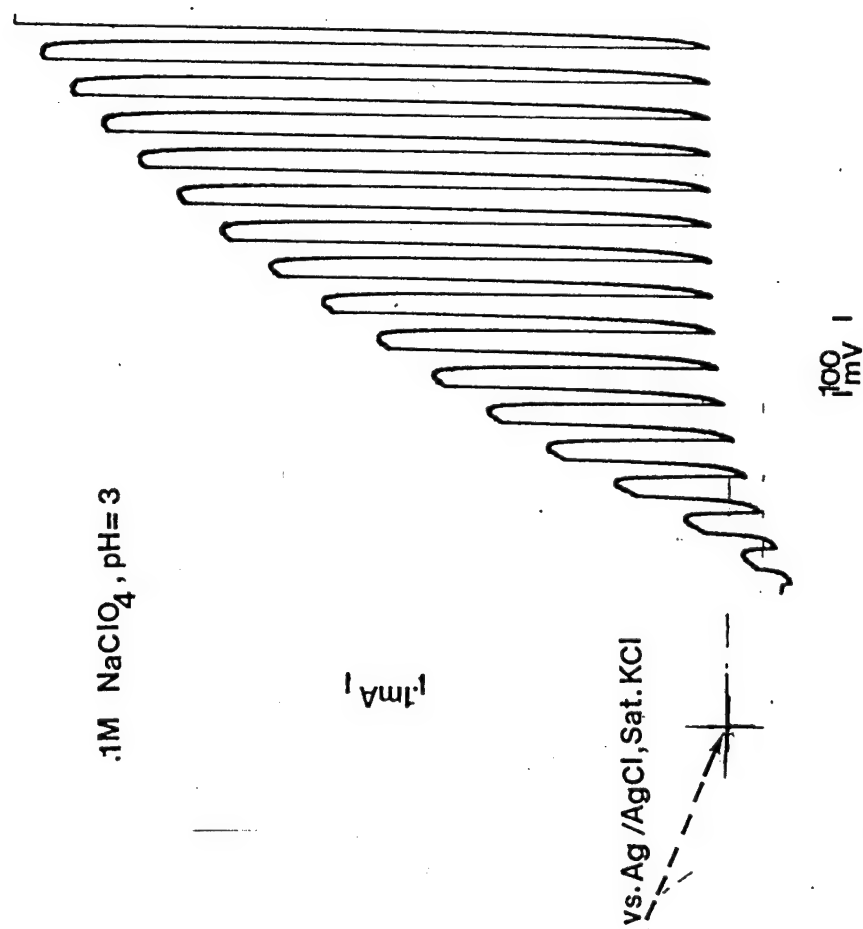
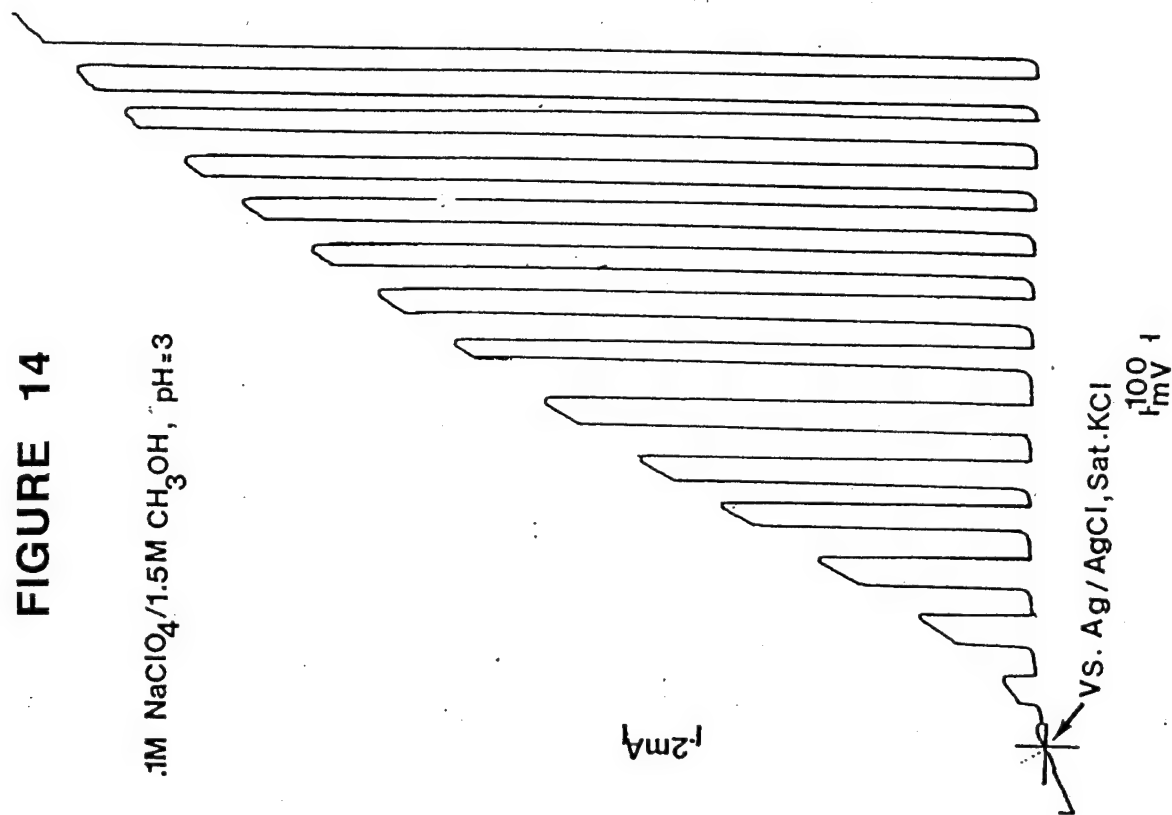


FIGURE 13

FIGURE 14

.1M NaClO<sub>4</sub>/1.5M CH<sub>3</sub>OH, pH=3

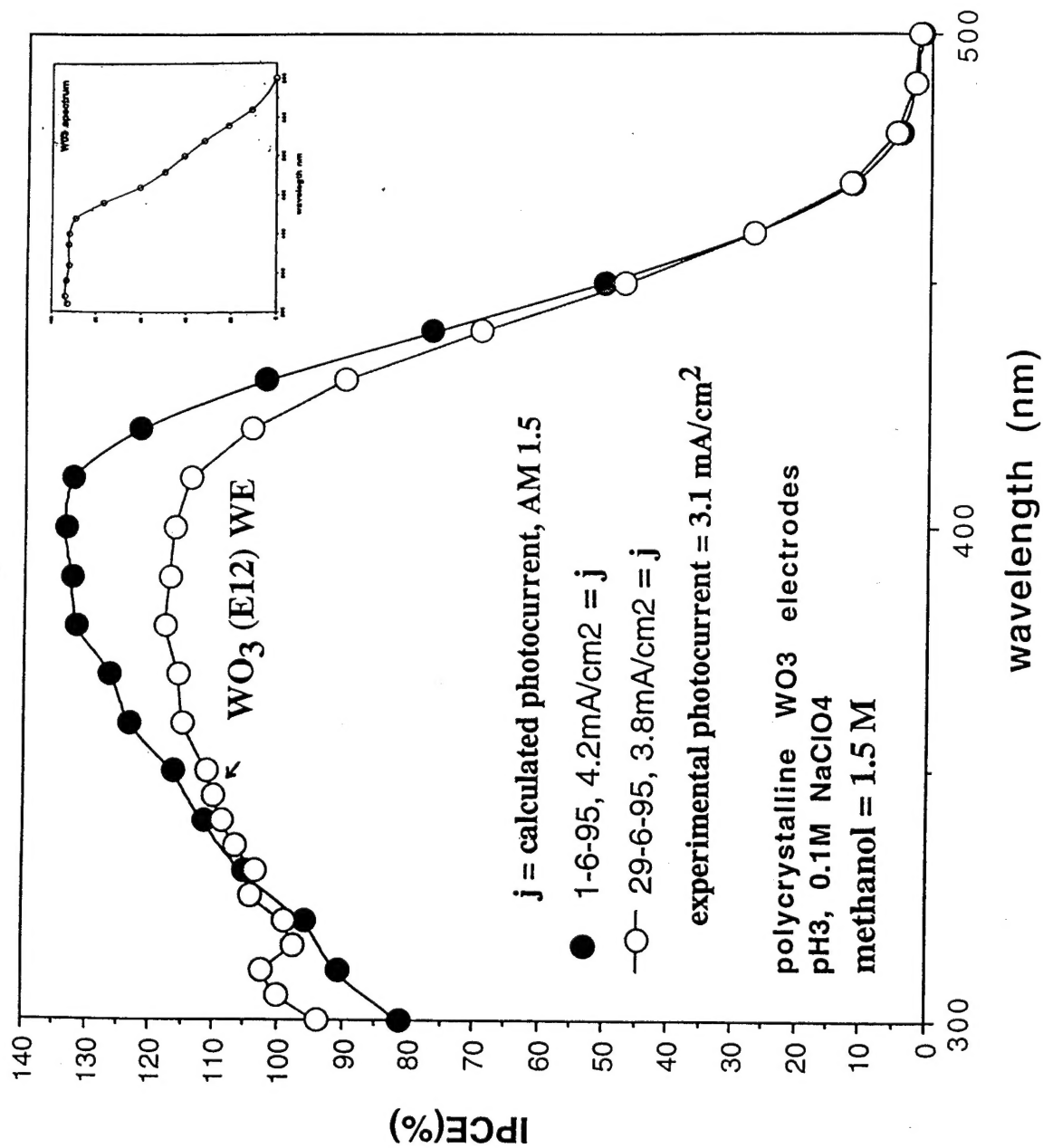


WHITE LIGHT PHOTOCURRENT/VOLTAGE PLOT  
FOR METHANOL OXIDATION USING WO<sub>3</sub> (E12) WE

wavel	dw	I(w)	I(w)xdw	$\Sigma I, 100\%$	$\Sigma I(w)xdw$	IPCE(%)	wave(nm)	wavel	$\Sigma I, 100\%$	IPCE(%)xdr	$\Sigma I, WOS$	$\Sigma WOS$
305.000	5.000	0.009	0.046	1.131	0.046	0.480933	490	305.000	1.131	63.33	0.72	0.72
310.000	5.000	0.041	0.204	6.231	0.250	0.598867	480	310.000	6.100	67.96	3.47	4.16
315.000	5.000	0.104	0.520	13.197	0.770	1.152248	470	315.000	13.197	71.27	9.41	13.59
320.000	5.000	0.174	0.872	22.503	1.642	2.661227	460	320.000	22.603	74.59	16.78	30.37
325.000	5.000	0.238	1.190	31.176	2.831	5.109492	450	325.000	31.176	79.54	24.80	55.17
330.000	5.000	0.361	1.905	60.698	4.736	9.398237	440	330.000	60.698	84.49	42.84	98.01
335.000	5.000	0.376	1.880	123.806	6.616	15.865033	430	335.000	60.790	88.66	45.03	143.04
340.000	5.000	0.420	2.098	174.598	8.714	26.091255	420	340.000	67.512	92.82	53.38	196.42
345.000	5.000	0.423	2.115	232.108	10.829	36.536144	410	345.000	67.512	93.37	54.94	251.36
350.000	7.500	0.466	3.497	290.953	14.325	49.013487	400	350.000	68.692	93.92	92.69	344.05
360.000	10.000	0.501	5.014	389.644	19.339	61.833908	390	360.000	145.568	93.39	135.94	479.99
370.000	10.000	0.642	6.421	535.212	25.760	74.801710	380	370.000	191.594	85.89	184.18	644.17
380.000	10.000	0.687	6.867	726.806	32.627	85.991699	370	380.000	210.440	74.80	187.41	801.58
390.000	10.000	0.695	6.946	937.247	39.373	93.385283	360	390.000	218.463	61.83	135.08	936.67
400.000	10.000	0.976	9.764	1156.710	49.337	93.918056	350	400.000	314.968	49.01	154.38	1091.04
410.000	10.000	1.116	11.162	1470.877	60.499	92.819836	340	410.000	369.096	36.54	134.64	1225.89
420.000	10.000	1.141	11.411	1839.744	71.910	84.494630	330	420.000	386.502	25.09	96.96	1322.66
430.000	10.000	1.033	10.330	2226.245	82.240	74.585740	320	430.000	368.218	15.87	66.83	1379.70
440.000	10.000	1.265	12.548	2584.463	94.788	67.982851	310	440.000	358.212	9.40	41.65	1421.54
450.000	10.000	1.471	14.707	3029.715	109.485	56.701532	300	450.000	446.252	5.11	27.27	1446.61
460.000	10.000	1.542	15.416	3563.436	124.911	54.830149	290	460.000	533.722	2.56	14.65	1463.46
470.000	10.000	1.524	15.237	4135.320	140.148	0.000000	280	470.000	571.894	1.15	6.65	1470.11
480.000	10.000	1.569	15.693	4712.652	155.841			480.000	577.631	0.80	3.64	1473.75
490.000	10.000	1.483	14.834	5320.323	170.876			490.000	607.471	0.49	2.88	1476.63
500.000	10.000	1.493	14.926	5906.505	185.801			500.000	607.471			
				6508.360	195.601				601.855			

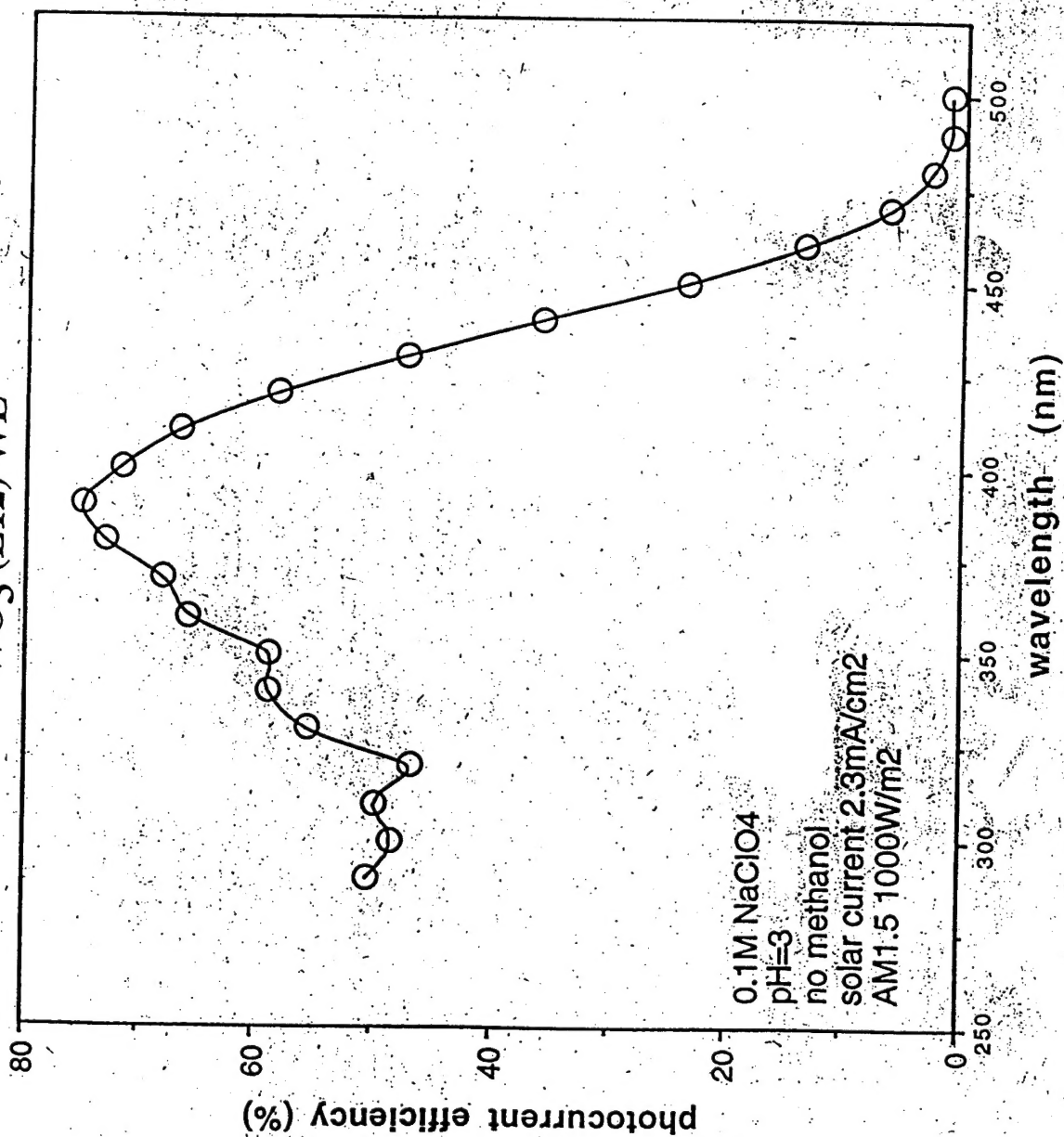
TABLE 1

# ACTION SPECTRA FOR METHANOL OXIDATION ON WO<sub>3</sub> FILMS



**FIGURE 15**

**ACTION SPECTRUM FOR WATER OXIDATION ON  
WO<sub>3</sub> (E12) WE**



**FIGURE 16**

# ACTION SPECTRUM FOR PHENOL OXIDATION ON TiO<sub>2</sub> FILMS

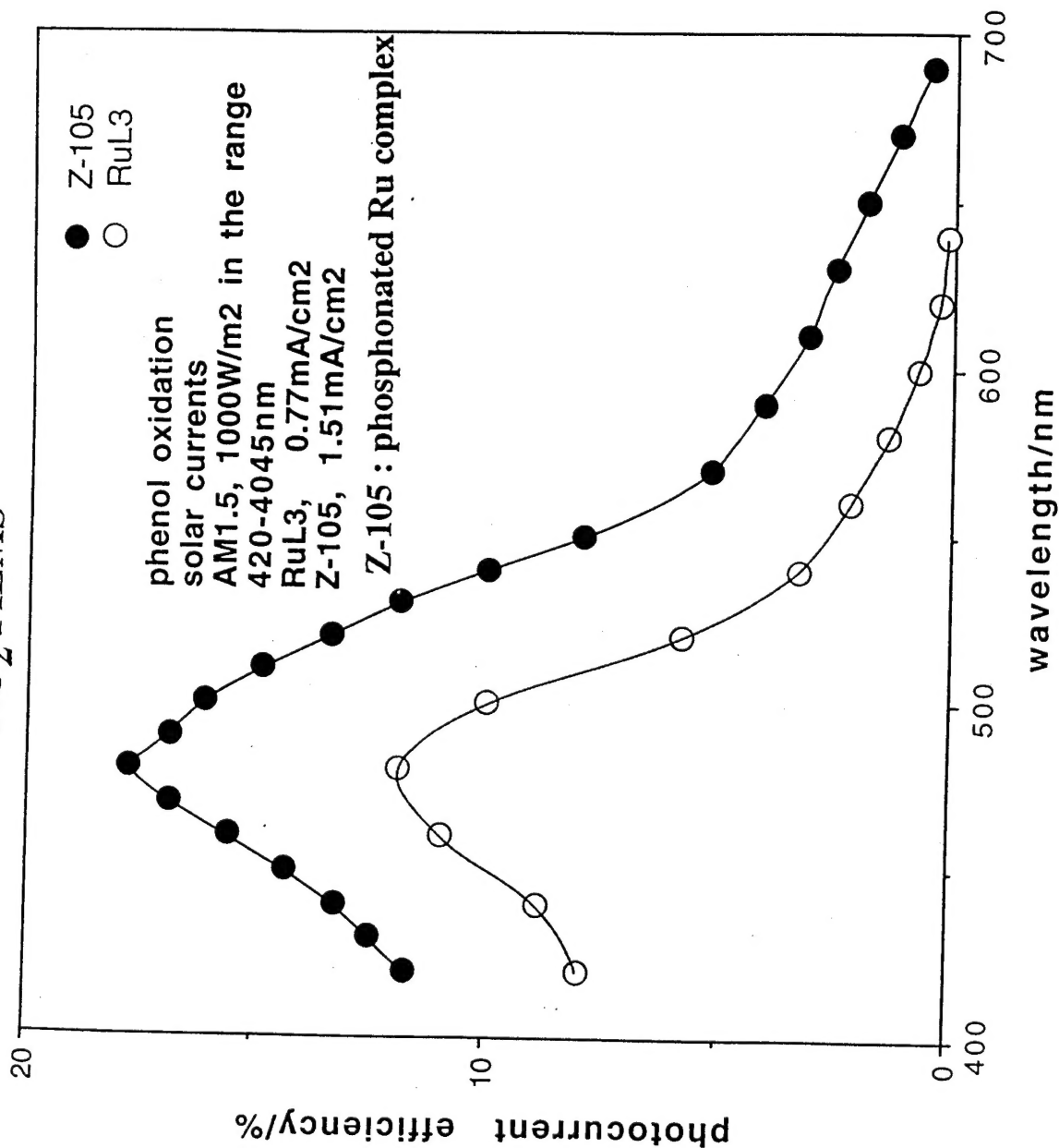


FIGURE 17